

**Roberto M. Alers Velázquez**  
University of Toledo  
1567 Twin Oaks Dr.  
Toledo, OH, 43615  
787-922-6104  
roberto.alers-velazquez@rockets.utoledo.edu

**EDUCATIONAL BACKGROUND:**

<b>University of Toledo</b> , Toledo, OH Doctor in Philosophy – Molecular Biology (In Progress) College of Natural Sciences and Mathematics PI: Dr. Scott Leisner	2015-Present
<b>University of Puerto Rico</b> , Aguadilla, PR Bachelor of Science Mayor in Biomedical Sciences Honors: Magna Cum Laude	2010 to 2014
<b>Colegio San Antonio</b> , Isabela, PR High School Diploma Honor Student	2010

**RESEARCH EXPERIENCE:**

<b>Ohio State University</b> <i>Project: Anthocyanin Biosynthesis pathway regulation</i> SiGuE Program- Research Fellow The Dorothy M. Davis Heart & Lung Research Institute Mentor: Dra. Andrea Doseff <i>Relevant Skills acquired:</i> DNA-Protein interaction assays Yeast transformations Transcription factor binding analysis	July 2014 – July 2015
-------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------	-----------------------

**University of Colorado - Anschutz Medical Campus**

Summer 2013

*Graduate Experience for Multicultural Students (GEMS)*

Department of Neurology, Aurora, CO

En Passant Mutagenesis was done on VZV to determine if ORFs 63/70 plays a role in virus replication.

Mentor: Dr. Ravi Mahalingam

*Relevant Skills acquired:*

Primer Design for Mutagenesis

Bacmid DNA extraction

*En Passant* Mutagenesis

Red Recombination

Transfection

PCR

Ligation

Gel extraction and DNA cleanup

**UPR Biology Department**

2012 - 2013

University of Puerto Rico, Aguadilla, PR

*Undergraduate Research- **Susceptibility of Halobacteria to Antibiotics***

Analysis of susceptibility of Halobacteria to certain antibiotics and determine the difference in behavior and which antibiotic is more effective toward the Halobacteria.

Mentor: Dr. Jose Planas

*Relevant Skills acquired:*

Preparation of Halobacteria Agar and Broth

Use of the Kirby-Bauer Test

Analyzing of culture behavior to the antibiotic

Serial-dilutions

Preparation of antibiotic solution for Kirby-Bauer test

**UPR Biology Department**

2010 – 2011

University of Puerto Rico, Aguadilla , P.R

*Undergraduate research- **Macroinvertebrates in Riparians wetland***

Studied the biodiversity of micro-invertebrates in a riparian wetland along the Guajataca River (Quebradillas, Puerto Rico) that was rich in micro-invertebrates.

We found a great number of Odonata individuals living among the large amount of the plant Chara that lurked in that area.

Mentor: Dr. Robert Mayer

*Relevant Skill acquired:*

Statistical analysis of invertebrates in certain markers

Classification of invertebrates

Use of the dissecting microscope

**University of Michigan Department of Biochemistry**

Summer 2012

University of Michigan, Ann Arbor, MI

*REU Summer undergraduate Research- College of Pharmacy*

FprA enzyme where reacted with NADPH and (4<sup>2</sup>R)-NADPH and (4<sup>2</sup>S)-NADPH to determine an Isotope effect to see if the enzyme is “sticky”.

Mentor: Dr. Bruce Palfey

*Relevant skills acquired:*

Poured agar plates with ampicillin

Lyophilization

Flash Freezing

Affinity chromatography to purify protein obtained

pH Adjustments

Cultured bacteria from a -80°C freezer

Poured agar plates with and without antibiotics

Induced protein expression using IPTG

Poured and ran SDS-PAGE gels

Learned to troubleshoot pipette malfunctions

Learned to make cells chemically competent

Used a spectrophotometer

Learned how to use a Stop-Flow Spectrophotometer

Experienced with coomassie staining

Synthesized Isotopes for Reaction- (4<sup>2</sup>R)-NADPH and (4<sup>2</sup>S)-NADPH

Learned how to disrupt cells using a sonicator

Learned Desalting techniques- Desalting column

Learned how to prepare a variety of buffers for reactions done in the research

Experienced with the preparation of anaerobic reactions using a tonometer

## **PRESENTATIONS AND CONFERENCES:**

### *Poster Presentations:*

#### *Annual Biomedical Research Conference for Minority Students*

“Is Varicella-Zoster Virus ORF 63/70 expression required for virus Replication?”

Nashville, TN

November 2013

#### *245th American Chemical Society National Meeting & Exposition.*

“Analysis of Isotope Effects in FprA to determine whether a Substrate is “Sticky”

New Orleans, LA

April 7-11, 2013

#### *Annual Biomedical Research Conference for Minority Student*

“Analysis of Isotope Effects in FprA to determine whether a Substrate is “Sticky”

San Jose, Ca

November 2012

“Analysis of Isotope Effects in FprA to determine whether a Substrate is “Sticky”

Research Experience for Undergraduates, Ann Arbor, MI

August 2012

#### *American Society of Plant Biologist - Midwestern Section Annual Meeting*

“Characterization of PALs Transcriptional Regulation in Maize”

Outstanding Poster Presentation – 1<sup>st</sup> place

Donald Danforth Plant Science Center, St. Louis, MO

March 21-22, 2015

### *Oral Presentations:*

“Is Varicella-Zoster Virus ORF 63/70 expression required for virus replication”

49<sup>th</sup> JTM/34<sup>rd</sup> PRISM Puerto Rico Louis Stokes Alliance For Minority Participation

University of Puerto Rico, Cayey, PR

March 29, 2014

“Is Varicella-Zoster Virus ORF 63/70 expression required for virus Replication?”

*245th American Chemical Society National Meeting & Exposition.*

*Dallas, TX*

*March-16, 2014*

“Is Varicella-Zoster Virus ORF 63/70 expression required for virus Replication?”

14<sup>th</sup> Undergraduate Research Symposium, University of Puerto Rico at Aguadilla

Aguadilla, PR

April-3, 2014

“Analysis of Isotope Effects in FprA to determine whether a Substrate is “Sticky”  
 13<sup>th</sup> Undergraduate Research Symposium, University of Puerto Rico at Aguadilla  
 Aguadilla, PR March -7, 2013

“Analysis of Isotope Effects in FprA to determine whether a Substrate is “Sticky”  
 48<sup>th</sup> JTM/33<sup>rd</sup> PRISM Puerto Rico Louis Stokes Alliance For Minority Participation  
 Universidad del Turabo , PR March – 9, 2013

“Is Varicella-Zoster Virus ORF 63/70 expression required for virus Replication?”  
 NIH – Grad. Experience for Multicultural Students (GEMS), Denver, CO August -7 2013  
 Presentation of Vida Marina at San Carlos school  
 “*Importance of Sand Dunes and Leatherback turtles*” February 2011  
 Aguadilla, PR

Presentation of Vida Marina to 4H club  
 “*Importance of Sand Dunes and Leatherback turtles*” March 2011  
 Fajardo, PR

Tourism Festival representing Vida Marina November 2011  
 Aguada, PR

Vida Marina Orientation in Open house at University of Puerto  
 Aguadilla, PR November 2010

Presentation to the FFA about Vida Marina Project  
 “*Importance of Sand Dunes and Leatherback turtles*” November 2010  
 San Sebastian, PR

## Professional Affiliations/Memberships

- American Chemical Society – Student Membership 2012-Present
- ACS Student Chapter , University of Puerto Rico, Aguadilla, PR 2012-2014
- ACS Student Chapter Green Chemistry Division-Assistant Director 2013-2014
- ACS Student Chapter Scientific Committee-Coordinator 2013-2014
- Caribbean Center for the Reduction of Aquatic Debris, Aguadilla, PR 2010-Present
- The Endocrine Society In-Training Associate Comp Member 2012-2013
- American Society for Microbiology 2013- Present

## RELEVANT COURSEWORK:

### University of Puerto Rico

Introduction to Biomedical Sciences	Social Psychology	Anatomy & Physiology II
General Biology I, II	Cell & Molecular Biology	Anatomy & Physiology Lab II
General Biology Lab I, II	Cell & Molecular Biology	Lab II
General Chemistry I, II	Physics I	Physics II
Gen. Chemistry Lab I, II	Physics I Lab	Statistics
Organic Chemistry I, II	Bioethics	Advanced Molecular Biology
Organic Chem. Lab I, II	Virology	Biology
Calculus I	Virology Lab	Research Methodology
Genetics	Immunology	
Genetics Lab	Immunology Lab	
Microbiology	Anatomy & Physiology I	
Microbiology Lab	Anatomy & Physiology Laboratory I	

## **ADDITIONAL SKILLS:**

### *Introductory courses and demonstrations concerning:*

Utilizing and preparing Agarose gel& SDS-PAGE

Scanning electron microscopy

Adobe Photoshop

Adobe Illustrator

KaleidaGraph

ChemDraw

PADI Certification:

Open Water Diver, Diver No. 12090G1235

### *Skills acquired from educational laboratory courses:*

Micropipetting

Aseptic technique

Pour plating, spread plating, and  
streak plating

Gel electrophoresis and imaging

Use of Image J analysis

Induction of competency in *E. coli*

PCR

Transformation of *E.coli*

Photospectroscopy

Restriction digests

Creation of dilution series

Gram-staining, spore staining,  
capsule staining, and simple staining

Bacterial plate screens and selection

Electroporation

Gateway Cloning

## **References:**

- Dr. Ravi Mahalingam – Contact information: (303) 724-4311/ [ravi.mahalingam@ucdenver.edu](mailto:ravi.mahalingam@ucdenver.edu)
- Dr. Bruce Palfey – Contact Information: (734) 615-2452/ [brupalf@umich.edu](mailto:brupalf@umich.edu)
- Dr. Carlos Ruiz-Martinez – Contact Information: (787) 319-5442/ [carlos.ruiz8@upr.edu](mailto:carlos.ruiz8@upr.edu)
- Dr. Jose Planas – Contact Information: (787) 646-5860/ [jose.planas@upr.edu](mailto:jose.planas@upr.edu)
- Dr. Scott Leisner- Contact information: [scott.leisner@utoledo.edu](mailto:scott.leisner@utoledo.edu)